REVIEW AND INTERPRETATION

Ecological importance of *Neotyphodium* spp. grass endophytes in agroecosystems

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Keywords

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Abstract

Neotyphodium spp. fungal endophytes form symbiotic associations with agronomic grasses with expression ranging from mutualistic to parasitic. In general, endophyte infection frequencies seem more variable in natural compared to pastoral situations, suggesting that expression and benefits attributable to endophyte infection depend on the resource environment in which the host-endophyte complex was formed and the conditions under which it is being grown. Emerging evidence suggests that expression is not always mutualistic or beneficial to the host in terms of productivity and persistence in resource-limited environments. Expression of host-endophyte associations will vary depending on resource availability and environmental conditions that influence host physiology and growth, and herbivore behavior. New research suggest alkaloids and secondary metabolites produced by host-endophyte associations can have physiological functions in addition to acting as herbivore deterrents. Novel host-endophyte associations are being created that maintain insect and disease resistance, while having minimal detrimental impact on mammalian herbivores. The influence of host-endophyte associations on sward composition and soil food webs is only now coming to light. While novel endophyte associations and their attendant mechanisms can be considered beneficial in some respect, the associations may not be as adaptable to stressful or marginal resource environments and could have long-term ecological impacts measured in terms of persistence and total productivity of the sward. We review the extensive published work on host-endophyte interactions and illustrate the complexity of host-endophyte associations and their interactions with environment, and the range of responses that occur.

Many grasses have evolved as symbiotic associations with fungi, including ecto- and endomycorrhizal fungi of the roots (Smith and Read, 1997), and fungi that systemically infect grass shoots. Among the latter, fungi that live their entire life cycle within the aerial portion of the host grass forming nonpathogenic, systemic, and usually intercellular associations are commonly defined as grass endophytes (Bacon and De Battista, 1991). These endophytes are classified in the tribe Balansieae of the family Clavicipitaceae (Ascomycetes) (Diehl, 1950). Within the past two decades, seven genera of grass endophytes were identified, including *Atkinsonella*, *Balansia*, *Balansiopsis*, *Echinodothis*, *Epichloë*, *Myriogenospora*

(White, 1994), and *Parepichloë* (White and Reddy, 1998). The best known grass endophytes are *Neotyphodium* spp. endophytes (e-endophytes), which are an anamorphic stage of *Epichloë* spp., endophytes: *N. coenophialum* (Morgan-Jones & Gams) Glenn, Bacon & Hanlin, found in tall fescue (*Lolium arundinaceum* [Schreb.] SJ Darbyshire, formerly *Festuca arundinacea* Schreb.), *N. uncinatum* (Gams, Petrini & Schmidt) Glenn, Bacon & Hanlin, colonizing meadow fescue (*L. pratense* [Huds.] SJ Darbyshire, formerly *F. pratensis* [Huds.]), and *N. lolii* (Latch, Christensen & Samuels) Glenn, Bacon & Hanlin, in perennial ryegrass (*L. perenne* L.). Their importance is related to the agronomic impact measured in terms of

Table 1 Similarities and differences between e-endophytes and p-endophytes, and grass host responses to infection

Property	e-endophytes	p-endophytes	Source	
Incidence	Pooideae (numerous)	Festuca, Lolium, Achnatherum	Siegel (1993)	
			An <i>et al</i> . (1993)	
			Hayes and Faeth (2004)	
			Li and Nan (2004)	
Mycelium in seed	Dense	Sparse	Siegel <i>et al.</i> (1984)	
			Siegel <i>et al.</i> (1995)	
Mycelium in leaf	Intercellular; nonbranched,	Inter- or intracellular, highly	Christensen et al. (2002)	
	synchronized growth	branched, unsynchronized growth		
Mycelium in roots	No	Yes	Hinton and Bacon (1985)	
			Siegel <i>et al.</i> (1995)	
Sporulation on host/Infectivity	No	Yes	An <i>et al</i> . (1993)	
Host tolerance to biotic	Yes	?	Schmidt (1993);	
and abiotic factors			Malinowski et al. (1997a)	
Production of alkaloids	Yes	?	Richardson (2000)	
			Siegel <i>et al.</i> (1995)	
Seed dissemination	Yes	Yes	Clay and Schardl (2002)	

livestock performance (Oliver, 2005) and sward persistence. Detailed studies with grasses led to the discovery of at least three other groups of symptomless fungal endophytes. One group (p-endophytes) consists of closely related Gliocladiumlike endophytes (An et al., 1993) in perennial ryegrass (Latch et al., 1984; Philipson, 1991) and Phialophora-like endophytes in meadow fescue, F. gigantea (L) Vill., and Arizona fescue (F. arizonica Vasey) (Schmidt, 1991; An et al., 1993). The pendophytes are ordered to Eurotiales (Ascomycetes) (Siegel et al., 1995) and are not related to e-endophytes (Leuchtmann, 1992). There are substantial differences between e-endophytes and p-endophytes not only in their morphology and physiology, but also in grass host responses to infection (Table 1). The third group of grass endophytes (a-endophytes) has been proposed to accommodate endophytes found in Italian ryegrass (L. multiflorum Lam.) and F. paniculata L. (Naffaa et al., 1998), and other annual species of the Lolium genus (Guillaumin et al., 2000). These endophytes are represented by parasitic species of Acremonium similar to A. chilense, an endophyte of orchardgrass (Dactylis glomerata L.) (Morgan-Jones et al., 1990). Similar to p-endophytes, effects of a-endophytes on physiological processes and ecology of cool-season grasses are not well understood at present.

Clay (1988) defined interactions between *Neotyphodium* spp. endophytes and cool-season grasses as defensive mutualism, where infected plants have generally higher resistance to a range of biotic and abiotic stresses (Malinowski and Belesky, 2000). Wilkinson and Schardl (1997) revised the concept and defined these interactions as asymptomatic symbioses. In such associations, the loss of sexuality is a fundamental cost in the long term in contrast to associations that have sexual and asexual means of reproduction. Detailed

studies, however, indicate that symbioses of grasses with Neotyphodium spp. endophytes can result both in antagonistic or mutualistic associations, suggesting a continuum from parasitism to mutualism (Saikkonen et al., 1998; Spyreas et al., 2001). It is not known how grass host and the endophyte recognize each other, nor how the grass host interacts at the molecular level to tolerate or eliminate the endophyte. There is circumstantial evidence that grass hosts respond to the presence of Neotyphodium spp. endophytes in a way that is similar to infection with pathogenic fungi, at least in part based on secondary metabolites the partners produce (Schulz et al., 1999). These interactions may result in a balanced antagonism and, consequently, coexistence of the symbiota (as in the case of Neotyphodium spp. endophytes) or an imbalanced antagonism leading to a disease of the host plant (as for pathogenic fungi) (Table 2).

We outline principal research findings and conclusions on ecological significance of *Neotyphodium* spp. endophytes for agronomic and nonagronomic grasses, and consider the agronomic significance and future application of novel endophytes in agroecosystems.

Primary driving forces in the evolution of *Neotyphodium*/grass associations

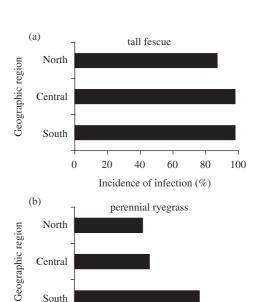
Greater tolerance to biotic stresses, such as protection from herbivores, nematodes, pathogenic fungi, and neighboring plants, could be primary forces in the evolution of *Neotyphodium*/grass associations, resulting in a dominance of endophyte-infected (E+) rather than noninfected (E-) grasses in native grasslands (Schardl and Phillips, 1997). Results obtained on tall fescue and perennial ryegrass pastures in the

Table 2	Responses of	arass hosts	to infection with	nathogenic fungi	and Neotyphodium-endophytes	
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Defense component	Pathogenic fungi	Neotyphodium-endophytes	Source
Chitinase	Yes	Yes	Roberts <i>et al.</i> (1992) Salzer <i>et al.</i> (2001)
Resveratrol	Yes	Yes	Sylvia and Sinclair (1983) Powell <i>et al</i> . (1994)
Other phenolic compounds	Yes	Yes	Malinowski <i>et al.</i> (1998a) Montesinos (2000)
Compatibility	No	Yes/No	Christensen (1995)
Effects on host plants	Imbalanced antagonism resulting in disease	Balanced antagonism resulting in coexistence	

USA and New Zealand seem to support this hypothesis (Popay and Bonos, 2005). Consider, however, that many agronomically important Festuca and Lolium species evolved in Europe as components of native grasslands, where grazing pressure was modest compared to those encountered in USA or New Zealand. Others propose more complex explanations for evolution of endophyte-grass host associations that involve multi-species interactions, multiple levels of causation and multidirectional flows of influence, which are modified by stochastic events, such as climate, soil and local biota (Saikkonen et al., 2004). Endophyte-infected cool-season grasses occur throughout continental and maritime temperate regions, generally with increased occurrence of endophyte infection detected at lower latitudes. It is quite likely, however, that increased concentrations of a suite of bioactive alkaloids would occur along with the increased infection frequency, because phytoalkaloids increase in numerous plant species as latitude decreases, from cool-temperate to tropical regions (Levin and York ,1978).

The incidence of E+ grasses in Europe may be correlated with climatic conditions (Lewis, 2000b), such that higher endophyte infection rate occurs in drier and hotter regions of southern Europe and lower endophyte infection rates in cool and moist environments of northern Europe (Figure 1). Recent findings suggest a close relationship between endophyte infection rate in grasses and adverse environment (Hahn et al., 2004; Sugawara et al., 2004). In fact, benefits of evolutionary adaptations to a specific environment in E+ compared to E- grasses may fail completely once host plants are exposed to contrasting conditions (Hesse et al., 2003). For example, E+ plants of perennial ryegrass ecotypes from a dry environment expressed reduced growth under wet conditions, but accelerated regrowth under drought stress when compared to E- plants. Endophyte-infected ecotypes of perennial ryegrass from dry and occasionally flooded environments produced more reproductive stems and seed, but fewer vegetative tillers when compared to E- plants. These responses are common adaptations of plants growing in drought-prone environments. Finally, endophyte infection in ecotypes of



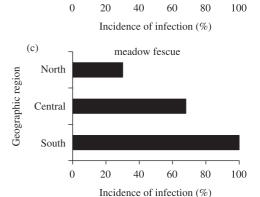


Figure 1 Incidence of endophyte infection in tall fescue, perennial ryegrass, and meadow fescue across precipitation and temperature gradients in Europe. The annual precipitation increases from south to north, while mean temperature decreases. Modified from Lewis *et al.* (1997) and Lewis (2000b).

perennial ryegrass originating in wet environments expressed heightened sensitivity to drought and reduced regrowth rates following drought. Specific host–endophyte associations may only be beneficial in the site of adaptation and can very likely be antagonistic outside of that environment.

Sleper (1985) proposed that tall fescue (2n = 6x = 42)evolved separately north and south of the Mediterranean Sea, as suggested by a lack of cross-compatibility between ecotypes collected in the two regions (Hunt and Sleper, 1981). DNA sequence analysis and fungal endophyte surveys helped identify the ranges of the two tall fescue groups: North Africa to the Western Mediterranean, Iberia and Italy, bounded at the Pyrenees and the Alps (the southern ecotypes), and northern Europe into Iberia, Morocco and Italy (the northern ecotypes) (Craven et al., 2005). Some of the southern ecotypes of tall fescue (from southern Spain, Algeria, and Sardinia) harbor endophytes (designated FaTG-2 and FaTG-3) that are genetically, biochemically, and morphologically different from N. coenophialum (Christensen et al., 1993; Tsai et al., 1994; Clement et al., 2001; Piano et al., 2005), whereas N. coenophialum is consistently found in northern tall fescue and in those tall fescue lineages introduced from northern Europe into North America, Australia and New Zealand (Craven et al., 2005). Interestingly, many recently developed cultivars of Mediterranean tall fescue (also referred to as summer-dormant types), such as Fraydo (Reed et al., 2004), Prosper (Heritage Seeds, 2005) or Flecha (Bonarino J, Gentos SA, Argentina, personal communication), do not harbor any endophytes suggesting their parental lines were also endophytefree. Moreover, artificial infection of Mediterranean-origin tall fescue with N. coenophialum does not seem to confer any benefits in terms of increased drought tolerance or persistence in hot and dry environments (Malinowski et al., 2005a; Malinowski, unpubl. data, but see Assuero et al., 2000). Thus, extended range of adaptation of tall fescue to drier and hotter environments, as a result of the association with N. coenophialum, may be important for the northern ecotypes but not plant ecotypes originating in hot and dry environments.

Conquering hot and dry environments

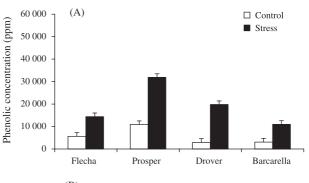
If the hypothesis that *Neotyphodium* spp. endophytes primarily enabled cool-season grasses to colonize drier and hotter environments is true, then the ability to survive drought should be commonly expressed by all cool-season grass hosts harboring these endophytes. This would most likely occur by developing efficient mechanisms improving water uptake, conserving water in plant tissues, or protecting meristematic tissues from oxidative stress. Earlier research identified positive effects of the shoot-located *Neotyphodium* spp. endophytes on a range of characteristics increasing drought stress tolerance in the three major agronomic grasses: perennial ryegrass, tall fescue, and meadow fescue, and negligible or detrimental

effects at optimal soil water supply (Malinowski and Belesky, 2000). In effect, the proportion of E+ grasses rapidly increases in pastures within a few years (Clay, 1988; Hill et al., 1998). In typically semidry environments, native grass populations also express a very high percentage of E+ plants (Sabzalian et al., 2005). Some recent work proposes, although strictly hypothetically, the ecological significance of newly discovered associations of *Neotyphodium* spp. endophytes with nonagronomic cool-season grasses in terms of greater abiotic stress tolerance (Marshall et al., 1999; Vinton et al., 2001). Detailed research on ecology of E+ native grasses, however, is extremely scarce. Nan and Li (2001) conducted a survey of native grasses in China, reporting endophyte infection in 25 species of 13 genera. The authors observed significant increase in root and shoot DM, and tiller number in response to endophyte infection in Hordeum bogdanii (Wilensky) and Elymus cylindricus (Franch.). In Arizona fescue, a grass native to the south-western USA, endophyte infection increased plant fitness only if exposed to soil water deficit (Morse et al., 2002), similar to results obtained with grasses having agronomic utility. Under limited soil water conditions, E+ plants of Arizona fescue had greater above-ground biomass and relative growth rate (RGR) than E-plants that correlated with higher net assimilation rates (P_n) , lower leaf conductance (g1), and greater specific leaf area (production of thinner leaves). Regardless of soil water availability, E+ plants tended to have lower midday P, than E- plants, primarily due to stomatal response, rather than biochemical limitations to photosynthesis. The adaptations are very similar to those described in agronomic grasses such as tall fescue and meadow fescue (Malinowski and Belesky, 2000). In contrast, stomatal functions do not seem to be affected by infection with N. lolii in perennial ryegrass, and there are either inconsistent or negligible effects for both physiological (Barker et al., 1997; Amalric et al., 1999) and morphological traits (Cheplick et al., 2000; Lewis, 2000a).

One important adaptation of plants to drought is the expression of an extensive root system that allows water uptake from a greater volume of soil. More massive root systems were reported in E+ compared to E- plants of the three agronomic grasses: perennial ryegrass (Latch et al., 1985), tall fescue (De Battista et al., 1990) and meadow fescue (Malinowski et al., 1997b), and a nonagronomic, rangeland species with the common name of sleepygrass (Achnatherum robustum [Vasey] Barkworth) (Hayes and Faeth, 2004). Although Neotyphodium spp. endophytes have not been found in roots of host grasses, they do affect root morphology and functions. Malinowski et al. (1999b) observed smaller root diameter and longer root hairs in hydroponically grown E+ compared to E- genotypes of tall fescue. A similar response was later shown for perennial ryegrass (Zhou et al., 2003). This phenomenon was not observed in soil-grown plants to date; however, smaller root diameter and longer root hairs will increase root surface area and should improve the water and nutrient uptake efficiency by plants manifesting these characteristics. In Arizona fescue, endophyte infection did not affect root DM or root-shoot ratio, regardless of available soil moisture (Morse *et al.*, 2002). Root morphology was not quantified.

Secondary metabolites of host-endophyte associations could be involved in the osmotic response of host grasses to soil water deficit stress (Lyons et al., 1990; Richardson et al., 1992; Bush et al., 1993; Elbersen and West, 1996). One theory suggests that loline alkaloids are involved in the regulation of drought responses in E+ grasses (Malinowski and Belesky, 2000). Although we now know that loline alkaloids are produced by Neotyphodium spp. endophytes from the amino acid proline, an abundant and common stress metabolite of many plants (Blankenship et al., 2005), direct involvement of the pyrrolizidine alkaloids in drought stress responses has not been documented. Another theory suggests that antioxidants are involved in the superior drought survival of E+ grasses, possibly by increasing tolerance of oxidative stress at the cellular level (Malinowski et al., 2005a). Endophyte-infected grasses contain much higher concentrations of phenolic compounds than their E- counterparts (Powell et al., 1994; Malinowski et al., 1998a; Zhou et al., 2003). Phenolics are potent antioxidants and help protect plant cells from oxidative stress during drought (Blokhina et al., 2003). Endophyteinfected plants of tall fescue (Ayad, 1998; Fike et al., 2001) and perennial ryegrass (Anzhi et al., 2004) express greater activity of superoxide dismutase (SOD) than E- plants. Activity of another enzyme associated with oxidative stress, ascorbate peroxidase, was higher in E+ than in E- plants of perennial ryegrass exposed to zinc (Zn) stress, suggesting that N. loliiendophyte modifies plant metabolism by favoring H₂O₂ scavenging throughout the catalase process (Bonnet et al., 2000). Interestingly, ongoing experiments (Malinowski, unpubl. data) with summer-dormant, Mediterranean cultivars of tall fescue (naturally free of endophytes) and orchardgrass showed greater concentrations of phenolic compounds in tiller bases when compared to summer-active cultivars of these grass species at the beginning of summer, regardless of soil moisture status (Figure 2). Thus, meristem survival during drought may be a function of protection from oxidative stress in summerdormant cultivars of a grass species. Protection could be amplified by endophytes in summer-active cultivars of the species. Another group of metabolites common to E-summerdormant (Volaire, 2002; Volaire et al., 2005) and E+ summeractive cool-season grasses (Carson et al., 2004) appears to be dehydrins, which apparently are correlated with the early sensing of soil water deficit and initiation of dormancy processes. The hypothesis implicating dehydrins in droughtstress tolerance of E+ grasses is worthy of further detailed investigation.

Neotyphodium spp. endophytes in seeds of agronomically important grasses are susceptible to heat damage especially



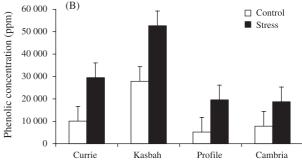


Figure 2 Phenolic concentrations in tiller bases of controlled (90% soil water capacity) and stressed (gradually imposed soil water deficit from 90% to 15% soil water capacity within 30 days) cultivars of naturally endophyte free tall fescue (A) and orchardgrass (B). For tall fescue, cultivars Flecha and Prosper are summer-dormant (Mediterranean), and Drover and Barcarella are summer-active. For orchardgrass, cultivars Currie and Kasbah are summer-dormant, and Profile and Cambria are summer-active. Bars indicate 1 s.e. (Malinowski, unpubl. data).

during long periods of storage (Siegel et al., 1984; Welty et al., 1987). Temperatures above 36°C are common in dry environments of southern Europe or northern Africa during summer. Hume and Barker (2005) reviewed data on endophyte survival in buried seed that were in contrast to findings on endophyte viability in storage systems. Interestingly, endophyte viability in seeds was higher in drier soil conditions. In the hot and dry environments of Tunisia, Morocco and Italy, endophyte infection frequencies in seeds of some indigenous tall fescue ecotypes were surprisingly high (Clement et al., 2001). We are not aware of any study specifically designed to evaluate endophyte survival rates in buried seeds under natural hot and dry environments; however, there seems to be no apparent problems with vertical transmission of endophytes.

Soil-root interface and soil microbial processes

Plants developed numerous adaptative strategies to deal with mineral imbalances, ranging from mineral deficiency to excess (Marschner, 1995). Such adaptations may enable certain species to dominate natural plant communities, especially where mineral imbalances occur. Traditionally, the ecological importance of grass endophytes was discussed in terms of drought tolerance and protection from herbivory. Recently, significant effort was devoted to understand interactions between leaf-located Neotyphodium spp. endophytes and root functions of cool-season grasses (Malinowski and Belesky, 2000). Mineral nutrients such as nitrogen (N) and phosphorus (P) were found to be associated with alkaloid production in E+ grasses (Belesky et al., 1988; Arechevaleta, 1992; Azevedo et al., 1993). One important mechanism of N assimilation discovered in E+ grasses was greater activity of glutamine synthetase, an enzyme responsible for NH₄ reassimilation (Lyons et al., 1990). This mechanism helped explain the efficient use of N by E+ tall fescue plants (Arachevaleta et al., 1989). A milestone step in our understanding of endophyte involvement in mineral nutrition was the discovery of chemical modifications in the rhizosphere of tall fescue and the regulatory effects root exudates had on uptake of certain minerals (Malinowski et al., 1998a; Malinowski and Belesky, 1999a, 1999b; Malinowski et al., 2000; Malinowski, et al., 2004). Release of phenolic-like compounds with iron (Fe3+) reducing and aluminum (Al) and copper (Cu) chelating activities facilitates P acquisition by E+ tall fescue plants (Malinowski and Belesky, 2000). Although roots of E+ perennial ryegrass plants also release phenolic compounds (Zhou et al., 2003), not much is known about their involvement in mineral uptake (Malinowski et al., 2004). Rahman and Saiga (2005) evaluated growth of E+ and E- clones selected from native populations of tall fescue in Japan cultivated on Black Andisol (low P content, high contents of other nutrients) and Red Andisol (high P content, low contents of other nutrients) under controlled growth conditions. When grown in Black Andisol, E+ plants had higher cumulative shoot dry weight and daily regrowth rates than E- plants, while in Red Andisol the reverse was true. In the P-deficient Black Andisol, endophyte infection increased P, potassium (K), calcium (Ca) and magnesium (Mg) uptake and transport within plants. Although root exudates were not evaluated in this study, results support the theory of an internal signaling system among the endophyte, grass host, and environment that leads to faster and more accurate responses to environmental factors than those occurring in E-plants.

A few recent observations (Monnet *et al.*, 2001, 2005) suggest greater tolerance of E+ compared to E− plants of perennial ryegrass to Zn. The response was measured in terms of damage to PSII system. The E+ plants were able to maintain photosynthesis and growth functions at higher Zn levels than were E− plants. The chelating abilities of phenolic compounds may be another mechanism enabling symbiotic grasses to cope with phytotoxic metals.

Soil microbial communities are linked with the plant community, the interaction of which may influence biodiversity

and ecosystem functioning (van der Heijden et al., 1998). Emerging studies show that endophytes have some influence on soil microbial processes, suggesting that E+ grasses not only have adaptative advantages in adverse environments, but can modify soil environments to ensure some competitive advantages (Matthews and Clay, 2001). Franzluebbers et al. (1999) demonstrated that tall fescue pastures containing a high percentage of E+ plants had greater soil organic C and N concentrations and lower soil microbial activity than pastures with low occurrence of endophyte infection. One explanation might involve endophyte metabolites, such as alkaloids and phenolics produced by the endophyte-grass host association, either actively released into the rhizosphere (Malinowski et al., 1998a, 1998b) or passively released from dead plant material (Franzluebbers et al., 1999). Support for the hypothesis of reduced microbial activity in soils grown with E+ Italian ryegrass was presented by Omacini et al. (2004), where leaf litter from E+ Italian ryegrass decomposed more slowly than E-leaf litter.

Grasses commonly form symbiotic associations with mycorrhizal fungi that increase grass host tolerance to drought and mineral deficiency stress (Miller, 1987; Smith and Read, 1997). Neotyphodium spp. endophytes can suppress mycorrhizal fungi colonization of grass roots in tall fescue (Baker, 1987; Chu-Chou et al., 1992; Guo et al., 1992), perennial ryegrass (Müller, 2003) and annual ryegrass (Omacini et al., 2005), suggesting alternative strategies to ensure plant survival under suboptimal environmental conditions. The ecological impact of this antagonistic interaction for E+ grasses is not clear; however, as shown by Vicari et al. (2002) mycorrhizae may actually reduce insect resistance in E+ perennial ryegrass plants, thus affecting competitive ability against other plants in mixed-sward grasslands. The in vitro growth of several pathogenic fungi (i.e. Fusarium culmorum, Dreschlera dictyoides) was inhibited in the presence of N. lolii (Holzmann-Wirth et al., 2001); however, responsible inhibitors were not yet identified.

Competitive ability and the avoidance of herbivory

The occurrence of E+ grasses in grasslands and managed pastures has had a significant and varied role in grazing animal behavior and human events through time (White et al., 2003). An array of bioprotective alkaloids produced by E+ grass hosts contributes directly to the competitive ability of the associations under a range of growing and management conditions (Bush et al., 1997). Knowledge of interactions of endophytes with agronomic species, and more recently grasses in natural or unmanaged environments, continues to expand (Clay, 1998; Clay and Holah, 1999; Brem and Leuchtmann, 2002; Faeth et al., 2004). In general, endophyte infection frequencies seem more variable in natural compared

to managed ecosystems supporting domesticated grasses such as tall fescue and perennial ryegrass. Variability suggests that expression and benefits attributable to endophyte infection depend on the resource environment in which the hostendophyte complex was formed and where it now occurs. Abundant nutrient supply might obscure endophyte advantages, measured in terms of plant growth, in competitive situations, but in some cases appears to facilitate secondary metabolite production and confer herbivory avoidance. Effects of endophyte infection on grazing livestock include depressed voluntary intake (Howard et al., 1992; Parish et al., 2003), altered duration, timing and intensity of grazing events (Eerens et al., 1998; Seman et al., 1999), reduced endocrine function and metabolic homeostasis (Porter and Thompson 1992; Bouton et al., 2002) decreased productivity measured in terms of live weight gain (Hill et al., 2002; Nihsen et al., 2004) and milk production (Thompson et al., 2001), and impaired reproductive processes including reduced conception, increased spontaneous abortions, altered gestation interval duration in females, and reduced spermatogenesis, testicular growth, testosterone production, and libido in males (Porter and Thompson, 1992). The effects on reproduction and ultimately numbers of grazers could contribute to reduced grazing pressure on natural grassland ecosystems with substantial E+ plant populations, and minimize usefulness of infected plant resources in managed agricultural applications.

Grazer selectivity could increase the likelihood that Egrasses would be consumed and in effect, shift composition and nutritive value of the sward to one that is inhospitable to herbivores because of the greater proportion of E+ plants. While depressed live weight gain arising from altered grazing behavior confounds the utility of E+ tall fescue in pasture, endophyte does help perpetuate swards with substantial E+ grass components, providing some level of primary production (e.g. available herbage in pasture), while at the same time minimizing soil erosion. Mitigation of soil erosion and the transition from intensive cropping to pasture production systems in the south-eastern USA was a major factor in the widespread use of E+ Kentucky 31 tall fescue. The importance and impact of endophyte infection on plant persistence and livestock productivity was unknown when Kentucky 31 was being introduced and applied as soil conservation and plant cover resource. The cultivar was able to grow and persist in pastures in the south-eastern US where other cool-season forage grasses could not. Population selection pressures exerted by large grazing animals, ruminants in the case of E+ tall fescue and perennial ryegrass, occur simultaneously with pressures associated with insects and microbes, including plant pathogens and host genotype. We present observations on influences of endophyte infection on plant responses at the individual and pasture community scale, where resource patches and sward structure dictate herbage productivity and nutritive value.

Grazing livestock performance on pasture in the USA tend to be problematic in the southern and western portions of the zone of tall fescue adaptation, because heat and drought stress influence the host-endophyte association as well as the grazer. Certain manifestations of the tall fescue toxicosis syndrome are weather-related with some aspects occurring in summer when grazer numbers on pasture typically are high and environmental conditions stress grazers. Elevated air temperatures reduced feed intake by cattle grazing E+ tall fescue, especially where temperatures exceeded 31°C (Hemken et al., 1984). Reduced herbage intake was caused by impaired ability to dissipate heat because of distal vasoconstriction (Rhodes et al., 1991). High temperature and restricted water availability increased alkaloid accumulation in E+ tall fescue plants (West, 1994), so that increased concentrations of bioactive alkaloids compounded the effects of extreme weather conditions on the grazer. Manifestations of the toxicosis syndrome, generally termed 'fescue foot' associated with distal vasoconstriction, can occur when cattle consume stockpiled E+ tall fescue in colder periods of the year. Alkaloid-induced physiological responses apparently increase grazer sensitivity to environmental extremes.

It seems that abiotic factors associated with regional and local climate, influence the host–endophyte-grazer complex, and can do so at many levels. For example, interactions could occur as:

- Host–endophyte genotype (plant and fungal genome expression when growing as symbionts with expression ranging from mutualism to antagonism; components could include native and novel nonergogenic endophytes, and high carbohydrate plant hosts)
- Host–endophyte genotype × soil resources (mineral nutrients, pH, water)
- Host–endophyte genotype × grazer genotype (breed and within breed variation; e.g. *Bos taurus* compared to *B. indicus*)
- Host–endophyte genotype × grazer genotype × abiotic environment (soil physical and biochemical limitations and climate associated with extremes in the range of plant adaptation)

All of the interactions are likely to be affected by extrinsic factors associated with management of agroecosystems and might not always be expressed in the same manner or to the same degree. For example, environmental conditions may stimulate plant host growth exceeding that of the endophyte within. This is a fairly typical response because the endophyte, being noninvasive, requires the intercellular space in which to grow and logically then plant growth should precede that of the endophytic mutualist.

There seems to be some difference of opinion regarding the nature of endophyte infection depending on whether the host plants are part of managed agroecosystems, or range and native grasslands. Endophyte associations with Arizona fescue are considered parasitic (Faeth and Sullivan, 2003), whereas others (Saikkonen *et al.*, 1998; Spyreas *et al.*, 2001)

suggest that a continuum of expression occurs between the symbionts ranging from parasitic to mutualistic. In their experiments, endophyte infection of tall fescue in intensively defoliated plots was associated with increased sward diversity, which they attributed to increased drought tolerance of E+ plants. It could be that mowing favored the infected plants that had accelerated leaf extension thereby increasing as a sward component through resource consolidation and competitive ability along the lines hypothesized by Barthram et al. (2005). In Barthram's experiments, resistance to invasive neighbors was related to residual sward surface height and the extension of new or existing leaves into space (to capture light). Because sward residue height was inversely related to resistance to invasion, the increased species richness observed by Spyreas et al. (2001) could be a product of leaf expression as well as increased water stress tolerance. In the case of tall fescue, sward residue height was directly related to alkaloid concentrations (Belesky and Hill, 1997) with probable impacts on alkaloid content and resistance to herbivory. Leaf elongation depended on host genotype especially early on during seedling growth (Belesky et al., 1989). In some instances, E- plants continue to grow in the presence of abiotic stresses (for instance water deficit), whereas E+ plants could adjust to stress by minimizing exposure to that particular stress (leaf rolling or slower growth compared to E- plants) and avoid physical and physiological damage.

Diverse communities seem to resist encroachment by other species especially where distribution of species is uniform rather than patchy (Tracy et al., 2004). Mutualistic endophytes such as those in tall fescue enhanced ability of the association to invade established plant communities, regardless of species composition of that community, although this is not always the case. Tracy and Renne (2005) observed that mixed species swards including an E-tall fescue cultivar, used to renovate a well-established tall fescue pasture, had few Eplants after 3 years. Shifts in sward composition could be a function of vigorous competition by E+ plants or preferential selection by grazers of plants other than E+ tall fescue. Rudgers et al. (2005) suggest that host-fungal associations may be more important early in the establishment phase than during the later growth phase of the sward. Changes in E+ grass litter and effects on mineral nutrient pools could influence the floristic composition of the sward and herbage productivity. Omacini et al. (2005) present an excellent synthesis of hostendophyte associations and ecosystem processes. A hierarchical relationship occurs at differing spatial and temporal scales. At the individual level, the endophyte influences individual host traits, affecting nutrient acquisition and interaction with other individuals in the sward and organisms at other trophic levels. This could influence structure of the sward (neighborhood) where abiotic stresses have some impact on productivity and persistence. At the community level, interaction with other species and resource distribution

patches occur. Endophyte infection affects nutrient pools, food web structure and ability to resist invasiveness, enabling E+ grasses to be more competitive in mixed species situations (Malinowski *et al.* 1999a).

A model (Faeth and Fagan, 2002) of the interaction of endophyte infection and stresses, predicts that low alkaloidproducing endophyte-grass associations should persist in population when soil nutrients and herbivory are low. Alternatively, high alkaloid endophyte-grass associations are favored under increasing herbivory and increasing soil nitrogen, at least to some point. At very high soil nitrogen levels, E- plants may be favored over either alkaloid-profile type of E+ plants. Predictions are supported by patterns of infection and alkaloid production in nature, and in field experiments; however, plant genotype and environmental factors, such as available water, interact with endophyte to influence host plant performance. The benefit of endophyte infection is less when nutrient and water availability is high, corroborating observations that endophyte facilitates host growth in stressful environments.

A host-endophyte association adapted to and arising from the environmental selection pressures of extreme conditions (e.g. high temperature and periodic and intense soil water deficits) and capable of producing alkaloids, might not express the same competitive fitness nor elicit the symptoms of fescue toxicosis in grazers when grown or used in a less stressful environment. Expression of toxicosis depends on the environment in which the association is grown, the nature of that association and management (e.g. livestock breed, nutrient inputs, grazing scheme) factors that influence the allocation and use of resources by the host-endophyte complex. Alkaloids accumulate under stressful conditions such as drought or when intermediate compounds, including numerous polyamines that can ameliorate the physiological impacts of water stress or serve as precursors to alkaloid synthesis, accumulate when plant growth is slow.

Domestication of *Neotyphodium*-endophytes

Recent shift from quantitative to qualitative assessment of mechanisms regulating adaptation of *Neotyphodium*—grass associations to abiotic stresses has resulted in domestication of the endophytes, that is, selection of novel endophyte strains that produce minimal amounts or no alkaloids detrimental to grazing livestock, yet appear to retain host tolerance to abiotic and biotic stresses (Tapper and Latch, 1999; Bouton *et al.*, 2002; West and Gunter, 2004). The major emphasis in selecting novel endophytes in the USA, Australia and New Zealand has been safety to grazing animals. Results from grazing experiments on Jesup tall fescue reinfected with MaxQ endophyte AR542 (Macoon *et al.*, 2004) and HiMag tall fescue reinfected with endophyte UA4 (Nihsen *et al.*,

2004), and perennial ryegrass reinfected with the Plus AR1 (New Zealand Agriseeds, 2005) or Plus Nea2 (Heritage Seeds, 2003) endophytes confirm excellent animal performance.

Ecological consequences of novel endophyte introduction into agroecosystems are not well understood. Research is currently underway to evaluate the persistence of grasses infected with novel endophytes in drought-prone environments. Bouton et al. (2002) observed about 15% decline in stand survival of summer-active Jesup MaxQ tall fescue when compared with Jesup infected with a wild-type endophyte. Both accessions survived much better than Jesup E- from 1997 through 2002 in the south-eastern USA. Tiller survival of Grasslands Flecha, a summer-dormant type tall fescue, infected with MaxQ endophyte AR542 was greater than that of E-plants during summer drought in the second and third growing seasons in a semiarid environment in North Texas, USA. Endophyte infection had no effect on tiller survival during an extreme summer drought at the same location in 2003 (Malinowski et al., 2005b). In the same study, neither AR542 nor native endophyte strains contributed to the persistence of summer-active tall fescue cultivars Jesup and Georgia 5 that did not survive the first summer drought. West and Gunter (2004) reported persistence of HiMag tall fescue infected with novel endophytes comparable with E+ Kentucky 31 and enhanced persistence relative to E-HiMag after 5 years of field trials in south-west Arkansas. Preliminary data (Malinowski, unpubl. data) do not confirm any advantages of the AR1 strain of N. lolii endophyte for drought tolerance in Aries and Quartet perennial ryegrass grown in a semiarid environment in North Texas, USA. In terms of mineral stress tolerance, the novel N. coenophialum strain, AR542, elicits similar responses in tall fescue host plants to those shown for the wild-type endophyte strains (Malinowski et al., 2004), i.e. exudation of phenolic-like compounds with metal chelating activity. Basic studies by Belesky et al. (2004) and Belesky and Burner (2004) suggest a slight disadvantage to juvenile plants of Jesup tall fescue of harboring the novel endophyte strain AR542 in terms of reduced germination and tillering rates when compared with plants infected with a native endophyte strain, or E- plants. Grasses infected with novel endophytes may experience a competitive disadvantage once exposed to stresses associated with defoliation and variation in growing conditions. Preliminary results, however, do not confirm any differences in terms of persistence of grasses that are endophyte free, or infected with either native or novel endophytes (Barker et al., 2005; Hume and Barker, 2005). One may argue that the time span in those studies was too short to make a comprehensive statement about plant persistence. For example, biotic stresses like insect herbivory (Popay and Baltus, 2001; Popay et al., 2004; Hunt and Newman, 2005) or nematode infestation (Timper and Bouton, 2004) were detrimental to grasses infected with novel endophytes when compared to grasses infected with native endophyte strains. Hunt and Newman (2005) suggest that AR542 infection does not provide the same degree of protection from certain insects (e.g. aphids) as does infection with the common strain endophyte. Repeated short-term disturbances of competitive ability might have drastic long-term consequences for stability of novel associations in managed agroecosystems. If similar results are found for other invertebrate herbivores, then the use of novel endophyte associations may be less useful in the wider sense than previously thought. The widespread conversion of tall fescue pastures, currently infected with common endophyte, to pastures composed of cultivars bearing the novel AR542 strain could lead to a general increase in the prevalence of certain insects and, potentially, diseases they spread. However, such conversion might be beneficial to native invertebrate herbivores, but not necessarily the host species.

References

Amalric C, Sallanon H, Monnet F, Hitmi A, Coudret A (1999)
Gas exchange and chlorophyll fluorescence in symbiotic and non-symbiotic ryegrass under water stress. *Photosynthetica* 37: 107–12.

An ZQ, Siegel MR, Hollin W, Tsai HF, Schmidt D, Bunge G, Schardl CL (1993) Relationships among non-*Acremonium* sp. fungal endophytes in five grass species. *Appl Environ Microbiol* 59: 1540–48.

Anzhi R, Yubao G, Yue C, Fang Z (2004) Effects of endophyte infection on POD, SOD, and PPO enzymes of perennial ryegrass (*Lolium perenne* L.) under drought stress. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 213.

Arachevaleta M, Bacon CW, Hoveland CS, Radcliffe DE (1989) Effect of the tall fescue endophyte on plant responses to environmental stress. *Agron J* 81: 83–90.

Arechevaleta M, Bacon CW, Plattner RD, Hoveland CS, Radcliffe DE (1992) Accumulation of ergopeptide alkaloids in symbiotic tall fescue grown under deficits of soil water and nitrogen fertilizer. *Appl Environ Microbiol* 58: 857–861.

Assuero SG, Matthew C, Latch CGM, Barker DJ, Haslett SJ (2000) Morphological and physiological effects of water deficit and endophyte infection on contrasting tall fescue cultivars. *N Z J Agric Res* 43: 49–61.

Ayad JY (1998) The effect of seaweed extract (*Ascophyllum nodosum*) on antioxidant activities and drought tolerance of tall fescue (*Festuca arundinacea* Schreb), Ph D Thesis, Texas Tech University, Lubbock, TX, USA, 158.

Azevedo MD, Welty RE, Creaig AM, Bartlett J (1993) Ergovaline distribution, total nitrogen and phosphorus content of two endophyte-infected tall fescue clones. In: *Proceedings of the 2nd International Symposium on Acremonium/Grass Interactions* (Eds Hume DE, Latch GCM, Easton HS), Palmerston North, New Zealand, 4–6 February 1993, AgResearch, Grassland Research Centre, Palmerston North, 59–62.

- Bacon CW, De Battista J (1991) Endophytic fungi of grasses. In: *Handbook of Applied Mycology*, Vol. 1. Soil and Plants (Eds Arora DK, Rai B, Mukerji KG, Knudsen GR), Marcel Dekker, New York, 231–256.
- Baker GM (1987) Mycorrhizal infections influences Acremonium-induced resistance to Argentine stem weevil in ryegrass. In: Proceedings of the 40th New Zealand Weed and Pest Control Conference, 11–13 August 1987. Nelson, New Zealand, 199–203.
- Barker DJ, Hume DE, Quigley PE (1997) Negligible physiological responses to water deficit in endophyte-infected and uninfected perennial ryegrass. In: *Neotyphodium/Grass Interactions* (Eds Bacon CW, Hill NS), Plenum Press, New York, 137–139.
- Barker DJ, Sulc RM, Bultemeier TL *et al.* (2005) Contrasting toxic-endophyte contamination between endophyte-free and nontoxic-endophyte tall fescue pastures. *Crop Sci* 45: 616–625.
- Barthram GT, Elston DA, Mullins CE (2005) The physical resistance of grass patches to invasion. *Plant Ecol* 176: 79–85.
- Belesky DP, Burner DM (2004) Germination and seedling development of a tall fescue cultivar in response to native and novel endophyte. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 305.
- Belesky DP, Burner DM, Ruckle JM (2004) Herbage mass and allocation of tall fescue as a function of native or novel, non-ergoalkaloid producing endophyte. In: *Proceedings of 2004 American Forage and Grasslands Council Conference* (Ed. Cassida K), 12–16 June 2004, Roanoke, VA, USA, 388–392.
- Belesky DP, Hill NS (1997) Defoliation and leaf age influence on ergot alkaloids in tall fescue. *Ann Bot* 79: 259–264.
- Belesky DP, Stringer WC, Hill NS (1989) Influence of endophyte and water regime upon tall fescue accessions. I. Growth characteristics. *Ann Bot* 63: 495–503.
- Belesky DP, Stuedemann JA, Plattner RD, Wilkinson SR (1988) Ergopeptine alkaloids in grazed tall fescue. *Agron J* 80: 209–212.
- Blankenship JD, Houseknecht JB, Pal S, Bush LP, Grossman RB, Schardl CL (2005) Biosynthetic precursors of fungal pyrrolizidines, the loline alkaloids. *Chembiochem* 6: 1016–1022.
- Blokhina O, Virolainen E, Fagerstedt KV (2003) Antioxidants, oxidative damage, and oxygen deprivation stress: a review. *Ann Bot* 91: 179–194.
- Bonnet M, Camares O, Veisseire P (2000) Effects of zinc and influence of *Acremonium lolii* on growth parameters, chlorophyll a fluorescence and antioxidant enzyme activities of ryegrass (*Lolium perenne* L. cv Apollo). *J Exp Bot* 51: 945–953.
- Bouton JH, Latch GCM, Hill NS *et al.* (2002) Reinfection of tall fescue cultivars with non-ergot alkaloid-producing endophytes. *Agron J* 94: 567–574.
- Brem D, Leuchtmann A (2002) Intraspecific competition of endophyte infected vs uninfected plants of two woodland grass species. *OIKOS* 96: 281–290.
- Bush LP, Fannin FF, Siegel MR, Dahlman DL, Burton HR (1993)

- Chemistry, occurrence and biological effects of saturated pyrrolizidine alkaloids associated with endophyte–grass interactions. *Agric Ecosyst Environ* 44: 81–102.
- Bush LP, Wilkinson HH, Schardl CL (1997) Bioprotective alkaloids of grass-fungal endophyte symbioses. *Plant Physiol* 114: 1–7.
- Carson RD, West CP, de los Reyes B, Rajguru S, Guerber CA (2004) Endophyte effects on dehydrin protein expression and membrane leakage in tall fescue. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 202.
- Cheplick GP, Perera A, Koulouris K (2000) Effect of drought on the growth of *Lolium perenne* genotypes with and without fungal endophytes. *Funct Ecol* 14: 657–667.
- Christensen MJ (1995) Variation in the ability of *Acremonium* endophytes of perennial rye-grass (*Lolium perenne*), tall fescue (*Festuca arundinacea*) and meadow fescue (*F. pratensis*) to form compatible associations in three grasses. *Mycol Res* 99: 466–470.
- Christensen MJ, Bennett RJ, Schmid J (2002) Growth of *Epichloe/ Neotyphodium* and p-endophytes in leaves of *Lolium* and *Festuca* grasses. *Mycol Res* 106: 93–106.
- Christensen MJ, Leuchtmann A, Rowan DD, Tapper BA (1993) Taxonomy of *Acremonium* endophytes of tall fescue (*Festuca arundinacea*), meadow fescue (*F. pratensis*), and perennial ryegrass (*Lolium perenne*). *Mycol Res* 97: 1083–1092.
- Chu-Chou M, Guo G, An ZQ *et al.* (1992) Suppression of mycorrhizal fungi in fescue by the *Acremonium coenophialum* endophyte. *Soil Biol Biochem* 24: 633–637.
- Clay K (1988) Fungal endophytes of grasses: a defensive mutualism between plants and fungi. *Ecology* 69: 10–16.
- Clay K (1998) Fungal endophyte infection and the population dynamics of grasses. In: *Population Biology of Grasses* (Ed. Cheplick GP). Cambridge University Press, Cambridge, UK, 255–285.
- Clay K, Holah J (1999) Fungal endophyte symbiosis and plant diversity in successional fields. *Science* 285: 1742–1744.
- Clay K, Schardl C (2002) Evolutionary origins and ecological consequences of endophyte symbiosis with grasses. *Am Naturalist* 160: S99–S127.
- Clement SL, Elberson LR, Youssef NN, Davitt CM, Doss RP (2001) Incidence and diversity of *Neotyphodium* fungal endophytes in tall fescue from Morocco, Tunisia, and Sardinia. *Crop Sci* 41: 570–576.
- Craven KD, Clay K, Schardl CL (2005) Systematics and morphology. In: *Tall Fescue on-Line Monograph Chapter 3* (Eds Fribourg HA, Hannaway DA), Oregon State University Extension Service and the University of Tennessee Agricultural Experiment Station, available online: http://forages.oregonstate.edu/is/tfis/monograph.html?PageID=124 [cited 21 October 2005].
- De Battista JP, Bouton JH, Bacon CW, Siegel MR (1990) Rhizome and herbage production of endophyte removed tall fescue clones and populations. *Agron J* 82: 651–654.

- Diehl WW (1950) *Balansia* and the Balansiae in America, USDA Agriculture Monograph No 4, Washington, DC, p 82.
- Eerens JPJ, Lucas RJ, Easton HS, White JGH (1998) Influence of the ryegrass endophyte (*Neotyphodium lolii*) in a cool-moist environment II. Sheep production. *NZJ Agric Res* 41: 191–199.
- Elbersen HW, West CP (1996) Growth and water relations of field-grown tall fescue as influenced by drought and endophyte. *Grass Forage Sci* 51: 333–342.
- Faeth SH, Fagan WF (2002) Fungal endophytes: Common host plant symbionts but uncommon mutualists. *Integ Comp Biol* 42: 360–368.
- Faeth SH, Helander ML, Saikkonen KT (2004) Asexual *Neotyphodium* endophytes in a native grass reduce competitive abilities. *Ecol Lett* 7: 304–313.
- Faeth SH, Sullivan TJ (2003) Mutualistic asexual endophytes in a native grass are usually parasitic. *Am Naturalist* 161: 310–325.
- Fike JH, Allen VG, Schmidt RE *et al.* (2001) Tasco-Forage. I. Influence of a seaweed extract on antioxidant activity in tall fescue and in ruminants. *J Anim Sci* 79: 1011–1021.
- Franzluebbers AJ, Nazih N, Stuedemann JA, Fuhrmann JJ, Schomberg HH, Hartel PG (1999) Soil carbon and nitrogen pools under low- and high-endophyte-infected tall fescue. *Soil Sci Soc Am J* 63: 1687–1694.
- Guillaumin J-J, Gendraud V, Naffaa W, Mouchacca J, Ravel C (2000) The a-endophytes, a new group of grass endophytes harboured by annual species of *Lolium*. In: *Proceedings of the 4th International Neotyphodium/Grass Interactions Symposium* (Eds Paul VH, Dapprich PD), 27–29 September 2000, Soest, Germany, 387–392.
- Guo BZ, Hendrix JW, An Z-Q, Ferris RS (1992) Role of *Acremonium* endophyte of fescue on inhibition of colonization and reproduction of mycorrhizal fungi. *Mycologia* 84: 882–885.
- Hahn H, Andreeva K, Wilner E, Diepenbrock W (2004) Fungal endophyte in grass species in Bulgaria and infection level in relation to altitude and water supply of their habitats. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 206.
- Hayes C, Faeth S (2004) Growth and competition of sleepy grass as influenced by co-occurring endophytes. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 312.
- van der Heijden MGA, Klironomos JN, Ursic M *et al.* (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* 396: 69–72.
- Hemken RW, Jackson JA Jr, Boling JA (1984) Toxic factors in tall fescue. *J Anim Sci* 58: 1011–1016.
- Heritage Seeds (2003) The Australian dairy pasture guide. 2003. Future prosperity ... New Grass, Heritage Seeds Pty Ltd., Melbourne, Australia, available online: http://www.heritageseeds.com.au/ASSETS/pdfs/brochures/2003DairyBro.pdf [cited 20 October 2005].
- Heritage Seeds (2005). Fescues, Heritage Seeds Pty Ltd,

- Melbourne, Australia, available online: http://heritageseeds.com.au/livestock/fescues.htm [cited 20 October 2005].
- Hesse U, Schöberlein W, Wittenmayer L *et al.* (2003) Effects of *Neotyphodium* endophytes on growth, reproduction and drought-stress tolerance of three *Lolium perenne* L. genotypes. *Grass Forage Sci* 58: 407–415.
- Hill NS, Belesky DP, Stringer WC (1998) Encroachment of endophyte-infected on endophyte-free tall fescue. Ann Bot 81: 483–488.
- Hill NS, Bouton JH, Thompson FN, Hawkins L, Hoveland CS, McCann MA (2002) Performance of tall fescue germplasms bred for high- and low-ergot alkaloids. *Crop Sci* 42: 518–523.
- Hinton DM, Bacon CW (1985) The distribution and ultrastructure of the endophyte of toxic tall fescue. *Canadian J Bot* 63: 36–42.
- Holzmann-Wirth A, Dapprich PD, Eierdanz S, Paul VH (2001) Anti-fungal substances extracted from *Neotyphodium* endophytes. In: *Proceedings of the 3rd International Conference* on Harmful and Beneficial Microorganisms in Grasslands, Pastures and Turf (Eds Paul VH, Dapprich PD), 26 September 2000, Soest, Germany, 65–69.
- Howard MD, Muntifering RB, Bradley NW, Mitchell GE, Lowry SR (1992) Voluntary intake and ingestive behavior of steers grazing Johnstone or endophyte-infected Kentucky-31 tall fescue. J Anim Sci 70: 1227–1237.
- Hume DE, Barker DJ (2005) Growth and management of endophytic grasses in pastoral agriculture. In: *Neotyphodium in Cool-Season Grasses* (Eds Roberts CA, West CP, Spiers DE), Blackwell Publishing Professional, Ames, Iowa, USA, 201–226.
- Hunt MG, Newman JA (2005) Reduced herbivore resistance from a novel grass–endophyte association. *J Appl Ecol* 42: 762–769.
- Hunt KL, Sleper DA (1981) Fertility of hybrids between two geographic races of tall fescue. *Crop Sci* 21: 400–404.
- Latch GCM, Christensen MJ, Samuels GJ (1984) Five endophytes of *Lolium* and *Festuca* in New Zealand. *Mycotaxon* 20: 535–550.
- Latch GCM, Hunt WF, Musgrave DR (1985) Endophytic fungi affect growth of perennial ryegrass. N Z J Agric Res 28: 165–168.
- Leuchtmann A (1992) Systematics, distribution, and host specificity of grass endophytes. *Natural Toxins* 1: 150–162.
- Levin DA, York BM Jr (1978) The toxicity of plant alkaloids: an ecogeographic perspective. *Biochem Syst Ecol* 6: 61–76.
- Lewis GC (2000a) Effect of drought stress on genotypes of *Lolium perenne* and other grass species with and without *Neotyphodium/Epichloë* infection. In: *Proceedings of the 4th International Neotyphodium/Grass Interactions Symposium* (Eds Paul VH, Dapprich PD), 27–29 September 2000, Soest, Germany, 201–205.
- Lewis GC (2000b) Neotyphodium endophytes: Incidence, diversity, and hosts in Europe. In: Proceedings of the 4th International Neotyphodium/Grass Interactions Symposium (Eds Paul VH, Dapprich PD), 27–29 September 2000, Soest, Germany, 123–130.
- Lewis GC, Ravel C, Naffaa W, Astier C, Charmet G (1997) Occurrence of *Acremonium* endophytes in wild populations

- of *Lolium* spp. in European countries and a relationship between level of infection and climate in France. *Ann Appl Biol* 130: 227–238.
- Li C, Nan Z (2004) A new *Neotyphodium* species symbiotic with drunken horse grass (*Achnatherum inebrians*) in China. *Mycotaxon* 90: 141–147.
- Lyons PC, Evans JJ, Bacon CW (1990) Effects of the fungal endophyte *Acremonium coenophialum* on nitrogen accumulation and metabolism in tall fescue. *Plant Physiol* 92: 726–732.
- Macoon B, Vann RC, Perkins JD III, Withers FT Jr (2004) Steer performance and forage production on novel-endophyte fescue compared to ryegrass pastures. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 504.
- Malinowski DP, Alloush GA, Belesky DP (1998a) Evidence for chemical changes on the root surface of tall fescue in response to infection with the fungal endophyte *Neotyphodium coenophialum*. *Plant Soil* 205: 1–12.
- Malinowski DP, Alloush GA, Belesky DP (2000) Leaf endophyte *Neotyphodium coenophialum* modifies mineral uptake in tall fescue. *Plant Soil* 227: 115–126.
- Malinowski DP, Belesky DP (1999a) Endophyte infection enhances the ability of tall fescue to utilize sparingly available phosphorus. *J Plant Nutr* 22: 835–853.
- Malinowski DP, Belesky DP (1999b) Infection with leaf fungal endophyte *Neotyphodium coenophialum* increases aluminum sequestration on root surfaces of tall fescue. *J Plant Nutr* 22: 1335–1349.
- Malinowski DP, Belesky DP (2000) Adaptations of endophyte-infected cool-season grasses to environmental stresses: Mechanisms of drought and mineral stress tolerance. *Crop Sci* 40: 923–940.
- Malinowski DP, Belesky DP, Fedders JM (1999a) Endophyte infection may affect the competitive ability of tall fescue grown with red clover. *J Agron Crop Sci* 183: 91–102.
- Malinowski DP, Belesky DP, Hill NS, Baligar VC, Fedders JM (1998b) Influence of phosphorus on the growth and ergot alkaloid content of *Neotyphodium coenophialum*-infected tall fescue (*Festuca arundinacea* Schreb). *Plant Soil* 198: 53–61.
- Malinowski DP, Belesky DP, Lewis GC (2005a) Abiotic stresses in endophytic grasses. In: *Neotyphodium in Cool-Season Grasses* (Eds Roberts CA, West CP, Spiers DE), Blackwell Publishing Professional, Ames, Iowa, USA, 187–199.
- Malinowski DP, Brauer DK, Belesky DP (1999b) *Neotyphodium coenophialum*-endophyte affects root morphology of tall fescue grown under phosphorus deficiency. *J Agron Crop Sci* 183: 53–60.
- Malinowski D, Leuchtmann A, Schmidt D, Nösberger J (1997a) Growth and water status in meadow fescue is affected by *Neotyphodium* and *Phialophora* species endophytes. *Agron J* 89: 673–678.
- Malinowski D, Leuchtmann A, Schmidt D, Nösberger J (1997b) Symbiosis with *Neotyphodium uncinatum* endophyte may

- increase the competitive ability of meadow fescue. *Agron J* 89: 833–839.
- Malinowski DP, Zuo H, Belesky DP, Alloush GA (2004) Evidence for copper binding by extracellular root exudates of tall fescue but not perennial ryegrass infected with *Neotyphodium* spp. endophytes. *Plant Soil* 267: 1–12.
- Malinowski DP, Zuo H, Kramp BA, Muir JP, Pinchak WE (2005b) Obligatory summer-dormant cool-season perennial grasses for semiarid environments of the southern Great Plains. *Agron J* 97: 147–154.
- Marschner H (1995) *Mineral Nutrition in Higher Plants*, 2nd edn. Academic Press Inc, Orlando, FL, USA, 889.
- Marshall D, Tunali B, Nelson LR (1999) Occurrence of fungal endophytes in species of wild *Triticum*. *Crop Sci* 39: 1507–1512.
- Matthews J, Clay K (2001) Influence of fungal endophyte infection on plant-soil feedback and community interactions. *Ecology* 82: 500–509.
- Miller RM (1987) The ecology of vesicular-arbuscular mycorrhizae in grass- and shrublands. In: *Ecophysiology of VA Mycorrhizal Plants* (Ed. Safir GR), CRC Press, Boca Raton, FL, USA, 135–170.
- Monnet F, Vaillant N, Hitmi A, Sallanon H (2005) Photosynthetic activity of *Lolium perenne* as a function of endophyte status and zinc nutrition. *Funct Plant Biol* 32: 131–139.
- Monnet F, Vaillant N, Vernay P, Coudret A, Sallanon H, Hitmi A (2001) Relationship between PSII activity, CO₂ fixation, and Zn, Mn and Mg contents of *Lolium perenne* under zinc stress. *J Plant Physiol* 158: 1137–1144.
- Montesinos E (2000) Pathogenic plant–microbe interactions. What we know and how we benefit. *Int Microbiol* 3: 69–70.
- Morgan-Jones G, White JF, Piontelli EL (1990) Endophyte–host associations in forage grasses. XIII. *Acremonium chilense*, an undescribed endophyte occurring in *Dactylis glomerata* in Chile. *Mycotaxon* 39: 441–454.
- Morse LJ, Day TA, Faeth SH (2002) Effect of *Neotyphodium* endophyte infection on growth and leaf gas exchange of Arizona fescue under contrasting water availability regimes. *Environ Exp Bot* 48: 257–268.
- Müller J (2003) Artificial infection by endophytes affects growth and mycorrhizal colonization of *Lolium perenne*. *Funct Plant Biol* 30: 419–424.
- Naffaa W, Ravel C, Guillaumin JJ (1998) A new group of endophytes in European grasses. *Ann Appl Biol* 132: 211–226.
- Nan ZB, Li CJ (2001) *Neotyphodium* in native grasses in China and observations on endophyte/host interactions. In: *Proceedings of the 4th International Neotyphodium/Grass Interactions Symposium* (Eds Paul VH, Dapprich PD), 27–29 September 2000, Soest, Germany, 41–50.
- New Zealand Agriseeds (2005) 2005 endophyte options. NZA Limited, Christchurch, New Zealand, available online: http://www.agriseeds.co.nz [cited 20 October 2005].
- Nihsen ME, Piper EL, West CP *et al.* (2004) Growth rate and physiology of steers grazing tall fescue inoculated with novel endophytes. *J Anim Sci* 82: 878–883.
- Oliver JW (2005) Pathophysiologic response to endophyte toxins.

- In: *Neotyphodium in Cool-Season Grasses* (Eds Roberts CA, West CP, Spiers DE), Blackwell Publishing Professional, Ames, Iowa, USA, 291–304.
- Omacini M, Chaneton EJ, Ghersa CM (2005) A hierarchical framework for understanding the ecosystem consequences of endophyte-grass symbioses. In: *Neotyphodium in Cool-Season Grasses* (Eds Roberts CA, West CP, Spiers DE), Blackwell Publishing Professional, Ames, Iowa, USA, 141–162.
- Omacini M, Chaneton EJ, Ghersa CM, Otero P (2004) Do foliar endophytes affect grass litter decomposition? A microcosm approach using *Lolium multiflorum*. *Oikos* 104: 581–590.
- Parish JA, McCann MA, Watson RH et al. (2003) Use of nonergot alkaloid-producing endophytes for alleviating tall fescue toxicosis in stocker cattle. J Anim Sci 81: 2856–2868.
- Philipson MN (1991) Ultrastructure of the Gliocladium-like endophyte of perennial ryegrass (Lolium perenne L.). New Phytol 117: 271–280.
- Piano E, Bertoli FB, Romani M *et al.* (2005) Specificity of host–endophyte association in tall fescue populations from Sardinia, Italy. *Crop Sci* 45: 1456–1463.
- Popay AJ, Baltus JG (2001) Black beetle damage to perennial ryegrass infected with AR 1 endophyte. *Proc N Z Grassl Assoc* 63: 267–271.
- Popay AJ, Bonos SA (2005) Biotic responses in endophytic grasses. In: *Neotyphodium in Cool-Season Grasses* (Eds Roberts CA, West CP, Spiers DE), Blackwell Publishing Professional, Ames, Iowa, USA, 163–185.
- Popay AJ, Silvester WB, Gerard PJ (2004) Effect of different endophyte isolates in perennial ryegrass on a root aphid, *Aploneura lentisci*. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 317.
- Porter JK, Thompson FN Jr (1992) Effects of fescue toxicosis on reproduction in livestock. *J Anim Sci* 70: 1594–1603.
- Powell RG, TePaske MR, Plattner RD, White JF, Clement SL (1994) Isolation of resveratrol from *Festuca versuta* and evidence for the widespread occurrence of this stilbene in the Poaceae. *Phytochem* 35: 335–338.
- Rahman HH, Saiga S (2005) Endophytic fungi (*Neotyphodium coenophialum*) affect the growth and mineral uptake, transport and effciency ratios in tall fescue (*Festuca arundinacea*). *Plant Soil* 272: 163–171.
- Reed KFM, Lee CK, Jahufer MZZ, Anderson MW (2004) Fraydo tall fescue (*Festuca arundinacea* L.). *Aust J Exp Agric* 44: 955–957.
- Rhodes MT, Paterson JA, Kerley MS, Garner HE, Laughlin MH (1991) Reduced blood flow to peripheral and core body tissues in sheep and cattle induced by endophyte-infected tall fescue. *J Anim Sci* 69: 2033–2043.
- Richardson MD (2000) Alkaloids of endophyte-infected grasses: Defense chemicals or biological anomalies?. In: *Microbial Endophytes* (Eds Bacon CW, White JF Jr). Marcel Dekker, Inc., New York, NY, USA, 323–340.
- Richardson MD, Chapman GW Jr, Hoveland CS, Bacon CW

- (1992) Sugar alkohols in endophyte-infected tall fescue. *Crop Sci* 32: 1060–1061.
- Roberts CA, Marek SM, Niblack TL, Karr AL (1992) Parasitic *Meloidogyne* and mutualistic *Acremonium* increase chitinase in tall fescue. *J Chem Ecol* 18: 1107–1116.
- Rudgers JA, Mattingly WB, Koslow JM (2005) Mutualistic fungus promotes plant invasion into diverse communities. *Oecologia* 144: 463–471.
- Sabzalian MR, Mirlohi A, Khayyam Nekoui M, Sharif Nabi B (2005) Distribution of Iranian palatable *Bromus tomentellus* Boiss. harboring novel *Neotyphodium* fungal endophytes. In: *Proceedings of the 1st International Conference on Plant-Microbe Interactions: Endophytes and Biocontrol Agents*, 18–22 April 2005, Saariselka, Finland (in press).
- Saikkonen K, Faeth SH, Helander M, Sullivan TJ (1998) Fungal endophytes: a continuum of interactions with host plants. *Ann Rev Ecol Syst* 29: 319–343.
- Saikkonen K, Wäli P, Helander M, Faeth SH (2004) Evolution of endophyte-plant symbioses. Trends Plant Sci 9: 275–280.
- Salzer P, Wiemken A, Boller T (2001) Different roles of plant chitinases in mycorrhizae and pathogenic interactions. In: *Chitin Enzymology* (Ed. Muzzarelli RAA). Atec Edizioni, Grottammare, Italy, 31–40.
- Schardl CL, Phillips TD (1997) Protective grass endophytes: Where are they from and where are they going? *Plant Dis* 81: 430–437.
- Schmidt D (1991) Les endophytes de la fétuque des prés. *Rev Suisse Agric* 23: 369–375.
- Schmidt D (1993) Effects of Acremonium uncinatum and a Phialophora-like endophyte on vigour, insect and disease resistance of meadow fescue. In: Proceedings of the 2nd International Symposium on Acremonium/Grass Interactions (Eds Hume DE, Latch GCM, Easton HS), Palmerston North, New Zealand, 4–6 February 1993, AgResearch, Grassland Research Centre, Palmerston North, 185–187.
- Schulz B, Rommert AK, Dammann U, Aust HJ, Strack D (1999)
 The endophyte–host interaction: a balanced antagonism? *Mycol Res* 103: 1275–1283.
- Seman DH, Stuedemann JA, Hill NS (1999) Behavior of steers grazing monocultures and binary mixtures of alfalfa and tall fescue. *J Anim Sci* 77: 1402–1411.
- Siegel MR (1993) Acremonium endophytes: our current state of knowledge and future directions for research. Agric Ecosyst Environ 44: 301–321.
- Siegel MR, Schardl CL, Phillips TD (1995) Incidence and compatibility of nonclavicipitaceous fungal endophytes in *Festuca* and *Lolium grass* species. *Mycologia* 87: 196–202.
- Siegel MR, Varney DR, Johnson MC *et al.* (1984) A fungal endophyte of tall fescue: Evaluation of control methods. *Phytopathology* 74: 937–941.
- Sleper DA (1985) Breeding tall fescue. In: *Plant Breeding Reviews* (Ed. Janick J), Vol. 3. AVI Publishing Company Inc, Westport, CT, USA, 313–342.
- Smith SE, Read DJ (1997) *Mycorrhizal Symbiosis*, 2nd edn. Academic Press, San Diego, CA, USA, 605.

- Spyreas G, Gibson DJ, Middleton BA (2001) Effects of endophyte infection in tall fescue (*Festuca arundinacea*: Poaceae) on community diversity. *Int J Plant Sci* 162: 1237–1245.
- Sugawara K, Yamashita M, Inoue T et al. (2004) Neotyphodium endophytes in naturalized ryegrasses in Japan: What are they doing. In: Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 215.
- Sylvia DM, Sinclair WA (1983) Phenolic compounds and resistance to fungal pathogens induced in primary roots of Douglas-fir seedlings by the ectomycorrhizal fungus *Laccaria laccata*. *Phytopathology* 73: 390–397.
- Tapper BA, Latch GCM (1999) Selection against toxin production in endophyte-infected perennial ryegrass. In: Ryegrass Endophyte: an Essential New Zealand Symbiosis. Grassland Research and Practice Series 7 (Eds Woodfield DR, Matthew C), New Zealand Grassl Assoc, Palmerston North, New Zealand, 107–111.
- Thompson FN, Stuedemann JA, Hill NS (2001) Anti-quality factors associated with alkaloids in eastern temperate pasture. *J Range Manage* 54: 474–489.
- Timper P, Bouton JH (2004) Effect of endophyte status and tall fescue cultivar on reproduction of lesion and stubby-root nematodes. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 406.
- Tracy BF, Renne IJ (2005) Reinfestation of endophyte-infected tall fescue in renovated endophyte-free pastures under rotational stocking. *Agron J* 97: 1473–1477.
- Tracy BF, Renne IJ, Gerrish J, Sanderson MA (2004) Effects of plant diversity on invasion of weed species in experimental pasture communities. *Basic Appl Ecol* 5: 543–550.
- Tsai HF, Liu JS, Staben C *et al.* (1994) Evolutionary diversification of fungal endophytes of tall fescue grass by hybridization with *Epichloë* species. *Proc Natl Acad Sci* 91: 2542–2546.
- Vicari M, Hatcher PE, Ayresi PG (2002) Combined effect of foliar and mycorrhizal endophytes on an insect herbivore. *Ecology* 83: 2452–2464.
- Vinton MA, Kathol ES, Vogel KP, Hopkins AA (2001) Endophytic

- fungi in Canada wild rye in natural grasslands. *J Range Manage* 54: 390–395.
- Volaire F (2002) Drought survival, summer dormancy and dehydrin accumulation in contrasting cultivars of *Dactylis glomerata*. *Physiol Plant* 116: 42–51.
- Volaire F, Norton MR, Norton GM, Lelievre F (2005) Seasonal patterns of growth, dehydrins and water-soluble carbohydrates in genotypes of *Dactylis glomerata* varying in summer dormancy. *Ann Bot* 95: 981–990.
- Welty RE, Azevedo MD, Cooper TM (1987) Influence of moisture content, temperature, and length of storage on seed germination and survival of endophytic fungi in seeds of tall fescue and perennial ryegrass. *Phytopathology* 77: 893–900.
- West CP (1994) Physiology and drought tolerance of endophyte-infected grasses. In: *Biotechnology of Endophytic Fungi of Grasses* (Eds Bacon CW, White JF Jr), CRC Press, Boca Raton, FL, USA, 87–99.
- West CP, Gunter SA (2004) Persistence of Hi-Mag tall fescue inoculated with nontoxic endophytes. In: *Proceedings of the 5th International Symposium on Neotyphodium/Grass Interactions* (Eds Kallenbach R, Rosenkrans C Jr, Lock TR), 23–26 May 2004, Fayetteville, AR, USA, Poster # 518.
- White JF Jr (1994) Taxonomic relationships among the members of the Balansiae (Clavicipitales). In: *Biotechnology of Endophytic Fungi of Grasses* (Eds Bacon CW, White JF Jr). CRC Press, Boca Raton, FL, USA, 3–20.
- White JF Jr, Bacon CW, Hywel-Jones N, Spatafora JW (2003) Clavicipitalean Fungi (Eds White JF Jr et al.). Marcel Dekker, New York, 575.
- White JF Jr, Reddy PV (1998) Examination of structure and molecular phylogenetic relationships of some graminicolous symbionts in genera *Epichloë* and *Parepichloë*. *Mycologia* 90: 226–234.
- Wilkinson HH, Schardl CL (1997) The evolution of mutualism in grass—endophyte associations. In: *Neotyphodium/Grass Interactions* (Eds Bacon CW, Hill NS). Plenum Press, New York, NY, USA, 13–25.
- Zhou F, Gao YB, Ma WJ (2003) Effects of phosphorus deficiency on growth of perennial ryegrass–fungal endophyte symbiont and phenolic content in root. *Plant Physiol Comm* 39: 321–325.